



UNIVERSITI PUTRA MALAYSIA

**ENZYMATIC EXTRACTION AND MODIFICATION, AND FRYING
STABILITY OF MORINGA OLEIFERA SEED OIL**

ABDULKARIM SABO MOHAMMED.

FSTM 2005 1

**ENZYMATIC EXTRACTION AND MODIFICATION, AND FRYING
STABILITY OF *Moringa oleifera* SEED OIL**

By

ABDULKARIM SABO MOHAMMED

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

January 2006



DEDICATIONS

This piece of work is dedicated to my parents, wife and children.

Abstract of the thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirements for the Degree of Doctor of Philosophy

**ENZYMATIC EXTRACTION AND MODIFICATION, AND FRYING
STABILITY OF *Moringa oleifera* SEED OIL**

By

ABDULKARIM SABO MOHAMMED

January 2006

Chairman: Professor Hasanah Mohd. Ghazali, PhD

Faculty: Food Science and Technology

Consumption of edible oils has grown with the increase in world population. The increasing health awareness and consciousness amongst consumers made the food industry more discriminating in the types of oil they use for food applications. Many circumstances have focused attention on high-oleic vegetable oils, which have been demonstrated to reduce the risk of coronary heart disease. The demand for high-oleic oils is increasing but there are only a few known sources available. *Moringa oleifera* seed oil, which is naturally high-oleic oil, therefore, presents a great opportunity for the oil industry for meeting this ever-increasing demand.

The objectives of this study were to determine the properties of oil extracted from *Moringa oleifera* seeds, evaluate the efficiency of enzymatic-extraction of the oil and modification of the oil to enhance its oleic acid content and compare the oxidative stability of the oil against several other oils during deep fat frying.

The oil content of *M. oleifera* seeds in Malaysia ranged between 30.8% and 33.4% depending on the variety, of which there were two. The physico-chemical properties of the oil were determined following extraction with light petroleum ether. The dominant fatty acid (FA) of the oil was indeed oleic acid, where Variety 1 contained 67.9% while Variety 2 contained 74.4%. After refining, the oil from both varieties is light golden color (0.1R + 1.0Y), and a viscosity, smoke point and refractive index ($n_D^{40^\circ\text{C}}$) of Cp 51.7, 206°C, and 1.4533, respectively. Using electronic nose analysis, the crude oil was found to have an odor similar to that of peanut oil. It has a complete melting point of 18.9°C. The crude oil contains 95.6% triacylglycerols (TAG) and 1.9% 1,2- and 1,3-diacylglycerols. The relative TAG content increased to 98.7% after refining. The oil contains 36.7% trioleoyl glycerol (OOO) as the main TAG.

M. oleifera seed oil was extracted using four different types of enzymes namely; Neutrase 0.8L (neutral protease), Termamyl 120L, type L (α -amylase), Pectinex Ultra SP-L (pectinase) and Celluclast 1.5L FG (cellulase) all supplied by Novozymes (Bagsvaerd Denmark). The enzymes were used either separately or in combination. The efficiency of enzyme-extraction was compared to aqueous extraction without enzyme. Enzymatic-extraction of *M. oleifera* seed oil showed that Neutrase alone at 2% v/w, 45°C and pH 6.8 was able to extract 71.9% oil relative to the amount obtained when the oil was solvent-extracted. Neutrase was the most efficient among the enzymes used followed by Termamyl, Celluclast and Pectinex with percent oil recoveries of 64.8%, 62.6% and 56.5%, respectively. Each extraction was carried out at the optimum pH and temperature of the enzymes. A combination of the four enzymes at pH 7.5 increased the

oil recovery to 74%. Percent oil recovery with all enzymes was significantly ($P<0.05$) higher than the control (aqueous extraction without enzyme) (35.6%).

Solvent extracted *M. oleifera* seed oil was transesterified using immobilized lipase (Lipozyme IM 60) (Novozymes Bagsvaerd Denmark) in order to change its melting and crystallizing behavior that will make it easier to fractionate. After transesterification, the oil was fractionated with acetone at -18°C and without acetone at 10°C to obtain two fractions, stearin and olein fractions. Incubation of the transesterified oil at 10°C for 24 h resulted in the formation of fat crystals, which settled at the bottom of the flask in sample transesterified for 24 h, while the control (0 h) sample became rather viscous with fat crystals in suspension. Transesterification affect the TAG profile of the oil, which in turn affected the solid fat content (SFC) and thermal behavior. The SFC value at 0°C after 24 h of reaction was 10.35% and significantly ($P<0.05$) higher than the control (0 h) (7.94%). The oil remained liquid at 20°C for all reaction times. The end set temperature (melting point) shifted from 18.9°C for the unreacted oil to 20.5°C for oil transesterified for 24 h. Transesterification of the oil resulted also in a significant ($P<0.05$) increase in the crystallization temperature of the high melting glyceride from the original value of 1.6°C to 12.9°C after transesterification for 24 h. There was a significant increase in the oleic acid content in the olein fractions obtained following fractionation of the transesterified oil with and without using acetone (75.2 and 70.5%, respectively) compared to the unreacted oil (67.9%).

The oxidative stability of refined *M. oleifera* seed oil (MoO) in deep fat frying was evaluated and compared with canola (CLO), soybean (SBO), and palm olein (PO). The oils were used to fry potato chips for 6 h a day up to a maximum of 5 days. Changes in fatty acid (FA) composition, free fatty acids (FFA), iodine value (IV) peroxide value (PV), *p*-anisidine value (*p*-AV), specific extinction ($E_{1\text{cm}}^{1\%}$ 233 and 269 nm for conjugated dienes and trienes), total polar compounds (TPC), color and viscosities were used to evaluate the oils.

The frying process caused an increase in the FFA contents MoO, PO, CLO and SBO. The FFA contents at the end of the frying period were 0.35%, 0.55%, 0.54% and 0.51% for CLO, PO, SBO and MoO, respectively. The rate of increase in the PV (meqO₂/kg) for CLO (2.33 per day) was higher compared to those of MoO (0.80 per day), PO (1.00 per day), and SBO (0.70 per day). Conjugated dienes levels at the end of the frying period were lowest in PO (4.27) followed by MoO (6.07) with high levels in CLO (9.28) and SBO (10.64). The amount of TPC in MoO (20.78%) and PO (21.23%) were significantly ($P < 0.05$) lower than those in CLO (28.73%) and SBO (31.82%). Color and viscosity of the oils increased with frying time. The rates of change of viscosity with the frying days were similar for all the oils. Results of sensory analysis conducted on potato chips fried in PO and MoO showed general acceptability of potato chips fried in both oils with high scores for crispness (7.07 and 7.14), oiliness (6.86 and 7.09), and fried food flavor (7.00 and 6.79) attributes, respectively. The overall acceptance of the French fries fried in MoO was high (7.50) and not significantly ($P > 0.05$) from that of PO (7.58).

Abstrak tesis ini dikemukakan kepada Senat Universiti Putra Malaysia bagi memenuhi keperluan untuk ijazah Doktor Falsafah.

**PENGKSTRAKAN BERENZIM, PENGUBAHSUAIAN BERENZIM DAN
KESTABILAN PENGGORENGAN MINYAK BIJI *Moringa oleifera*.**

Oleh

ABDULKARIM SABO MOHAMMED

Januari 2006

Pengerusi: Profesor Hasanah Mohd. Ghazali, PhD

Fakulti: Sains Makanan Dan Teknologi

Penggunaan minyak masak telah berkembang seiring dengan peningkatan bilangan penduduk dunia. Peningkatan tahap kesedaran terhadap kesihatan di kalangan pengguna menjadikan industri makanan semakin memilih tentang jenis minyak yang digunakan untuk aplikasi makanan. Banyak keadaan telah menumpukan perhatian kepada minyak sayuran tinggi asid oleik di mana minyak begini telah dibuktikan dapat mengurangkan risiko penyakit koronari jantung. Permintaan minyak tinggi asid oleik yang semakin meningkat tetapi sumbernya adalah terhad. Secara semulajadinya, minyak biji *M. oleifera* adalah minyak tinggi asid oleik, dan ini memberi peluang yang cerah kepada industri minyak untuk memenuhi permintaan ini.

Objektif kajian yang dijalankan ini adalah untuk menentukan ciri-ciri minyak yang diekstrak dari biji *Moringa oleifera*, menilai keberkesanan pengekstrakan minyak secara berenzim dan pengubahsuaian minyak tersebut secara berenzim untuk meningkatkan

kandungan asid oleik, dan membandingkan kestabilan pengoksidaan minyak tersebut dengan beberapa minyak lain semasa penggorengan dalam minyak penuh.

Minyak biji *M. oleifera* diekstrak menggunakan empat jenis enzim yang berbeza. Pengekstrakan berenzim telah dibandingkan dengan pengekstrakan tanpa enzim melalui menggunakan empat jenis enzim iaitu Neutrase 0.8L (protease neutral), Termamyl 120L, type L (α -amylase), Pectinex Ultra SP-L (pectinase) dan Celluclast 1.5L FG (cellulase) yang dibekalkan oleh Novozymes (Bagsvaerd Denmark). Enzim ini digunakan sama ada secara tunggal atau digabungkan. Kandungan minyak biji *M. oleifera* di Malaysia didapati dalam julat 30.8% dan 33.4%, bergantung kepada jenis varieti, di mana terdapat dua jenis varieti. Ciri-ciri fisiko-kimia minyak ditentukan setelah pengekstrakan menggunakan petroleum eter. Asid lemak paling utama adalah asid oleik, di mana Varieti 1 mempunyai sebanyak 67.9%, sementara Varieti 2 mempunyai sebanyak 74.4% asid oleik. Selepas penyulingan, minyak dari kedua-dua jenis varieti ini mempunyai warna cerah keemasan (0.1R + 1.0Y) dan nilai kelikatan, takat wasap dan indeks pembiasan iaitu ($n_D^{40^\circ\text{C}}$), Cp 51.7, 206°C dan 1.4533, masing-masing. Dengan menggunakan analisis hidung elektronik, minyak mentah didapati mempunyai bau yang seakan sama dengan minyak kacang. Takat leburnya ialah 18.9°C. Minyak kasar tersebut mengandungi 95.6% triasilgliserol (TAG), 1.9% 1,2- dan 1,3-diasilgliserol. Kandungan relatif TAG meningkat kepada 98.7% selepas penyulingan. Minyak tersebut mengandungi 36.7% triolilolgliserol (OOO) sebagai TAG yang utama.

Pengekstrakan berenzim minyak biji *M. oleifera* menunjukkan bahawa Neutrased (protease neutral daripada Novozymes Bagsvaerd Denmark) adalah paling berkesan dengan 71.9% penghasilan minyak, diikuti dengan Termamyl (α -amylase) (64.8%) Celluclast (cellulase) (62.6%) dan Pectinex (pectinase) (56.5%) dengan setiap pengekstrakan dijalankan pada pH dan suhu yang optimum bagi ke semua enzim tersebut. Gabungan empat jenis enzim berkenaan adalah lebih berkesan daripada penggunaan enzim secara bersendirian dengan penghasilan sebanyak 74%. pH yang digunakan adalah optimum bagi Neutrased. Peratus penghasilan minyak dengan kesemua enzim tersebut signifikannya ($P < 0.05$) lebih tinggi berbanding kawalan (pengekstrakan akues tanpa enzim) (35.6%).

Minyak biji *M. oleifera* yang diekstrak dengan pelarut telah ditransesterifikasi menggunakan lipase tersekat-gerak (Lipozyme IM 60) dengan tujuan menukar pelakuan penghabluran yang memudahkannya untuk dipisahkan. Selepas ditransesterifikasi, minyak tersebut dipisahkan dengan aseton pada suhu -18°C dan tanpa aseton pada suhu 10°C untuk memperolehi dua pecahan; pecahan stearik dan olein. Pengeraman minyak teresterifikasi pada 10°C selama 24 jam menyebabkan pembentukan hablur-hablur lemak yang termendak di dasar kelalang sementara minyak kawalan (0 jam) pula menjadi agak likat dengan hablur-hablur lemak yang terampai. Transesterifikasi menjejaskan profil TAG minyak tersebut di mana ia turut menjejaskan nilai kandungan lemak pepejal (KCP) dan pelakuan haba. Nilai KCP pada 0°C selepas tindakbalas 24 jam adalah 10.35% dan adalah lebih tinggi ($P < 0.05$) daripada kawalan (0 jam) (7.94%). Minyak tersebut kekal cair pada 20°C pada keseluruhan masa tindakbalas. Suhu akhir

yang ditetapkan (takat lebur) meningkat daripada 18.9°C bagi minyak tanpa tindakbalas kepada 20.5°C bagi minyak yang telah diesterifikasikan selama 24 jam. Transesterifikasi minyak juga mengakibatkan peningkatan yang bererti ($P < 0.05$) ke atas suhu penghabluran bagi takat lebur tinggi gliserida daripada nilai asalnya iaitu 1.6°C kepada 12.9°C selepas transesterifikasi selama 24 jam. Berlaku pertambahan yang bererti ke atas kandungan asid oleik dalam pecahan olein yang diperolehi berikutan dengan pemisahan minyak yang diesterifikasi dengan dan tanpa aseton (75.2% dan 70.5%) berbanding dengan minyak asal yang tidak ditindakbalas.

Kestabilan pengoksidaan minyak biji *M. oleifera* yang telah dituliskan dalam penggorengan minyak penuh telah dinilai dan dibandingkan dengan minyak kanola (CLO), minyak kacang soya (SBO) dan minyak kelapa sawit (PO). Minyak-minyak tersebut digunakan untuk menggoreng kentang selama 6 jam sehari sehingga maksimum 5 hari. Perubahan komposisi asid lemak, asid lemak bebas, nilai iodin, peroksida (PV), *p*-anisidin (*p*-AV), nilai pelupusan spesifik ($E_{1\text{cm}}^{\%}$ 233 dan 269 nm), jumlah sebatian polar (TPC), warna dan kelikatan telah digunakan untuk menilai minyak tersebut.

Aktiviti pengeringan yang dijalankan telah menyebabkan peningkatan kandungan FFA bagi minyak MoO, PO, CLO dan SBO. Kandungan FFA pada akhir masa penggorengan masing-masing adalah 0.35%, 0.55%, 0.54%, dan 0.51% bagi CLO, PO, SBO, dan MoO. Kadar pertambahan dalam nilai PV (meqO_2/kg) bagi CLO (2.33 per hari) adalah lebih tinggi berbanding dengan minyak MoO (0.80 per hari), PO (1.00 per hari) dan SBO (0.70 per hari). Paras diene konjugat pada akhir masa penggorengan adalah paling

rendah dalam PO (4.27) diikuti dengan aras tertinggi bagi CLO (9.28) dan SBO (10.64). Jumlah TPC dalam MoO (20.78%) dan PO (21.23%) adalah lebih rendah secara bererti ($P < 0.05$) berbanding dengan CLO (28.78%) dan SBO (31.82%). Warna dan kelikatan bagi minyak tersebut juga meningkat dengan masa penggorengan. Kadar perubahan kelikatan dengan masa penggorengan adalah hampir sama dengan kadar ke semua minyak tersebut. Keputusan analisis sensori menunjukkan penerimaan keseluruhan terhadap kentang yang digoreng dalam MoO dan PO, masing-masing, memberi markah tinggi bagi ciri-ciri seperti kerangupan (7.07 dan 7.14), rasa minyak (6.86 dan 7.09), dan citarasa makanan bergoreng (7.00 dan 6.79). Penerimaan keseluruhan terhadap kentang goreng dalam MoO adalah tinggi (7.50) dan tidak bererti ($P > 0.05$) dengan PO (7.58).

ACKNOWLEDGEMENT

In the name of Allah most beneficent most merciful. All praise be to Allah for all the favors bestowed upon Mankind. I wish to start by expressing my sincere gratitude to my supervisor, Professor Hasanah Mohd Ghazali of the Department of Food Science, Faculty of Food Science and Technology for all the support and guidance she offered throughout the period of this study. She made it possible for me to fulfill my ambition of studying for this degree by providing all the support and for been there for me all the time throughout the study period. I am indeed very grateful to my research committee members, in the names of Associate Professors Dr. Lai Oi Ming of Bioprocess Technology Department, Faculty of Biotechnology and Biomolecular Sciences, Sharifah Kharidah Syed Muhammad of Food Science Department, Faculty of Food Science and Technology and Dr. Kamariah Long of Malaysian Agricultural Research and Development Institute for their untiring support and advises which made this research feasible.

I would like to thank the technical and administrative staff of the Faculty of Food Science and Technology and the research staff of MARDI for their assistance. I am very thankful to my fellow graduate students in enzyme technology laboratory for being very friendly and supportive during difficult times.



I wish to acknowledge the Malaysian government for the IRPA grant awarded to Professor Dr. Hasanah Mohd Ghazali, with which this research was conducted and University Putra Malaysia for giving me the opportunity to study for the PhD degree.

Lastly I would like to thank my parents, my wife Fatima and my little Ruqayya for their support and care at all times.



I certify that an examination committee met on 20th January 2006 to conduct the final examination of Abdulkarim Sabo Mohammed on his Doctor of Philosophy thesis entitled “Enzymatic extraction and modification, and frying stability of *Moringa oleifera* seed oil” in accordance with Universiti Pertanian Malaysia (higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The committee recommended that the candidate be awarded the relevant degree. Members of the Examination Committee are as follows:

Yaakob Che Man, PhD

Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Chairman)

Nazamid Saari, PhD

Associate Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Internal Examiner)

Tan Chin Ping, PhD

Lecturer
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Internal Examiner)

David B. Min, PhD

Professor
Faculty of Food Science and Technology
Ohio State University
(External Examiner)



HASANAH MOHD GHAZALI, PhD

Professor/ Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: **16 FEB 2006**

This thesis submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee are as follows:

Hasanah Mohd Ghazali, PhD

Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Chairman)

Lai Oi Ming, PhD

Associate Professor
Faculty of Biotechnology and Biomolecular Science
University Putra Malaysia
(Member)

Sharifah Kharidah Syed Muhammad, PhD

Associate Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Member)

Kamariah Long, PhD

Department of Biotechnology
Malaysian Agricultural Research Development Institute (MARDI)
(Member)



AINI IDERIS, PhD

Professor/ Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: **09 MAR 2006**

DECLARATION

I hereby declare that the thesis is based on my original work except for quotations and citations, which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at UPM or other institutions.



ABDULKARIM SABO MOHAMMED

Date: 10th February 2006

TABLE OF CONTENTS

	Page
DEDICATION	ii
ABSTRACT	iii
ABSTRAK	vii
ACKNOWLEDGEMENTS	xii
APPROVAL	xiv
DECLARATION	xvi
LIST OF TABLES	xx
LIST OF FIGURES	xxiii
LIST OF PLATES	xxv
LIST OF ABBREVIATIONS	xxvi
 CHAPTER	
 1 INTRODUCTION	 1
 2 LITERATURE REVIEW	 9
Sources of oils	9
Oil Market	11
Uses of <i>Moringa oleifera</i>	15
Processes in oil extraction	16
Pretreatment of Raw material	18
Extraction methods	19
Traditional (wet) method	20
Mechanical method	22
Solvent method	23
Enzymatic method	24
Factors affecting enzymatic treatment of oil seeds and fruit	26
Temperature	26
pH	27
Dilution ratio	28
Enzyme concentration and combination	31
Reaction time	31
Centrifugation and shaking speed	32
Oil modification processes	32
Fractionation	34
Interesterification	37
Chemical interesterification	39
Enzymatic interesterification	40
Mechanism of enzyme reaction	42
Deep fat frying	44
Fried foods	46



	Quality of frying oil	46
	Changes that occur in oils during heating and frying	47
	Physical changes	
	Chemical changes	
	Measurement of frying oil deterioration	53
3	CHARACTERIZATION OF <i>Moringa oleifera</i> SEED OIL	
	Introduction	57
	Materials and Methods	59
	Materials	59
	Methods	60
	Results and Discussion	74
	Proximate analysis of <i>Moringa oleifera</i> seed	74
	Physical properties of <i>Moringa oleifera</i> seed oil	75
	Chemical properties of <i>Moringa oleifera</i> seed oil	88
	Summary	97
4	ENZYMATIC EXTRACTION OF <i>Moringa oleifera</i> SEED OIL	
	Introduction	98
	Materials and Methods	100
	Materials	100
	Methods	100
	Results and Discussion	105
	Oil extraction	105
	Effect of enzyme type and concentration and reaction time on oil recovery	105
	Effect of incubation temperature on oil recovery	111
	Effect of shaking speed on oil recovery	113
	Effect of enzyme combination and pH on oil recovery	114
	Physico-chemical properties of extracted oil	115
	Summary	120
5	OLEIC ACID ENHANCEMENT OF <i>Moringa oleifera</i> SEED OIL BY ENZYMATIC TRANSESTERIFICATION AND FRACTIONATION	
	Introduction	121
	Materials and Methods	123
	Materials	123
	Methods	124
	Results and Discussion	127
	Effect of transesterification on crystallization temperature of glyceride species	127
	Effect of reaction time on melting and crystallization behavior of <i>Moringa oleifera</i> seed oil	130
	Effect of reaction time on solid fat content	132
	Fatty acid composition of fractionated enzyme-reacted oil	135

	Melting behavior and iodine value of fractions from fractionated <i>Moringa oleifera</i> seed oil	139
	Summary	142
6	FRYING STABILITY OF HIGH-OLEIC <i>Moringa oleifera</i> SEED OIL IN COMPARISON WITH OTHER VEGETABLE OILS	
	Introduction	144
	Materials and Methods	146
	Materials	146
	Methods	147
	Results and Discussion	151
	Changes in fatty acid (FA) composition and ratios	151
	Changes in free fatty acid (FFA)	154
	Changes in iodine value (IV)	157
	Changes in peroxide value (PV)	158
	Changes in <i>p</i> -anisidine value (<i>p</i> -AV)	160
	Total oxidation (TOTOX) value	161
	Changes in specific extinction ($E_{1\text{cm}}^{1\%}$ at 233 and 269 nm)	162
	Changes in total polar compounds (TPC)	164
	Changes in color and viscosity	166
	Sensory analysis	171
	Summary	174
7	SUMMARY, CONCLUSIONS AND RECOMMENDATIONS	
	Summary	177
	Conclusions and Recommendations	179
	BIBLIOGRAPHY	181
	APPENDIX	195
	BIODATA OF THE AUTHOR	196
	PAPER PUBLISHED FROM THIS THESIS	197



LIST OF TABLES

Table	Page
1 Oilseeds: oil content, yield and producing areas	10
2 Major oilseeds: world production (million metric tones)	12
3 Import of 10 major oilseeds in million metric tones (MMT) by countries/regions importing more than 1 MMT	13
4 Major oilseed export (million metric tones)	14
5 Percent oil yield increase during enzymatic-extraction of oil- bearing materials at various pretreatment conditions	29
6 Factors affecting frying oil decomposition	49
7a Summary of frying parameters used in the evaluation of frying oils for stability	55
7b Properties used by various researchers to evaluate the frying performance and stability of oils	56
8 Proximate composition of <i>Moringa oleifera</i> seed	76
9 Physical properties of <i>Moringa oleifera</i> seed oil	78
10 Melting behavior of <i>Moringa oleifera</i> seed oil using different scan rates	82
11 Chemical properties of <i>Moringa oleifera</i> seed oil	89
12 Relative percent composition of fatty acid in <i>Moringa oleifera</i> seed oil	92

13	Triacylglycerol composition of solvent extracted <i>Moringa oleifera</i> seed oil	95
14	Enzyme manufacturer's specifications	102
15	Effect of enzyme type on oil recovery from <i>M. oleifera</i> seed	107
16	Physical and chemical Properties of <i>Moringa oleifera</i> seed oil extracted using different methods	117
17	Fatty acid composition of <i>Moringa oleifera</i> seed oil extracted using various methods	119
18	Crystallization behavior of original and enzyme-reacted <i>Moringa oleifera</i> seed oil (MoO)	129
19	Fatty acid composition (%) of MoO fractionated with and without solvent	136
20	MUFA, PUFA, SFA and total percent-unsaturated fatty acid contents of original and solvent fractionated MoO compared with some commercially available high-oleic oils	137
21	Melting behavior of original, solvent fractionated Liquid and solid fractions of enzyme-reacted MoO	141
22	Changes in fatty acid (FA) composition of oils during frying	152
23	Changes in free fatty acid content and iodine value for oils during frying	155
24	Changes in PV, <i>p</i> -AV and TOTOX value in oils during frying	159
25	Changes in specific extinction and total polar compounds in oils during frying	165

26	Changes in color and viscosity of oils during frying	167
27	Sensory attributes of potato chips fried in <i>M. oleifera</i> seed oil and palm olein	172
28	Frequency table of hedonic data for French fries fried in <i>M.oleifera</i> seed oil and palm olein	173



LIST OF FIGURES

Figures		Page
1	Palm oil dry fractionation routes	37
2	Triacylglycerol formation during interesterification of SOL	39
3	Representation of the reaction in a 1,3 specific enzymatic Interesterification	42
4	Catalytic mechanism for lipase-catalyzed interesterification, showing the catalytic site containing aspartic/glutamic, histidine and serine residues	43
5	Physical and chemical reactions occurring during frying	49
6	Hydrolysis reaction in frying oils	51
7	Oxidation reaction in frying oils	52
8	Polymerization reaction in frying oils	53
9	Heating thermograms of refined and crude <i>M. oleifera</i> seed oil at 5°C and 100°C/min scan rates	81
10	Cooling profile of crude and refined <i>M. oleifera</i> seed oils at 5°C/min scan rate	83
11	Solid fat content of crude and refined <i>M. oleifera</i> seed oil	84
12	Electronic nose flavor profiles of <i>M. oleifera</i> seed and Peanut oils	86
13	Fatty acid profile of <i>M. oleifera</i> seed oil obtained by gas chromatography	93
14	Triacylglycerol profile of <i>M. oleifera</i> seed oil	96



15	Effect of enzyme type and concentration on oil recovery	110
16	Effect of incubation time on oil recovery	110
17	Effect of incubation temperature on oil recovery	112
18	Effect of shaking speed on oil recovery	114
19	Effect of pH on oil recovery	115
20	DSC Heating thermogram of MoO after transesterification using enzyme for (A) 0 h, (B) 4 h, (C) 8 h, (D) 12 h, and (E) 24 h	131
21	DSC cooling thermogram of MoO after transesterification using enzyme for (A) 0 h, (B) 4 h, (C) 8 h, (D) 12 h, and (E) 24 h	131
22	Changes in SFC (%) of MoO after transesterification at various reaction times	133
23	TAG profile of (A) unreacted (B) enzyme-reacted <i>M. oleifera</i> seed oil after 24 h.	134
24	DSC heating thermogram of fractions from solvent fractionated MoO	140
25	Relationships between $C_{18:1}/C_{18:2} + C_{18:3}$ and the initial mounts of $C_{16:0} + C_{18:0}$ in oils to TPC and TOTOX values during frying	170